

Laboratory Diagnosis of Tb

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*Strains of the live vaccine *Mycobacterium bovis* BCG, which are also used intra-vesically in the treatment of bladder cancer, can occasionally cause disease in patients who are immunocompromised.

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Primary Infection



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Initial infection in a person by MTBC organisms is termed primary tuberculosis, usually site of entry, usually the lung. Lymph nodes will also be infected at this stage (primary complex) The tuberculin skin test (TST, Mantoux test) becomes positive at 3-8 weeks after infection, and marks the development of cellular immunity and tissue hypersensitivity.

This test is useful in detecting latent disease.

Interferon gamma release assays are also available.

Primary Infection





- Bronchoalveolar lavage/bronchial washings
- Gastric washings (Not all labs accept these)
- Sterile site body fluids(CSF>6ml needed)
- Urine specimens (x3 EMU)
- Skin, bone, and tissue including post mortem specimens (ideally caseous, not in formalin)
- Pus or pus swabs (Mycobacteria stick to flocking, so swabs not ideal)
- Faecal samples (not recommended)

Primary Infection



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- Blood
- Bone Marrow
- Pus
- Sterile Fluids

Can all be added directly to a Tb Culture Bottle

Post Primary





Foci developing in the endothelium of blood vessels may rupture leading to disseminated or miliary tuberculosis.

Post Primary



Post-primary tuberculosis develops either as a result of reactivation of organisms in a 'healed' primary lesion or because of exogenous re-infection.

Post-primary tuberculosis usually occurs five or more years after the primary infection and may affect children as well as adults. Infection with *M. tuberculosis* only progresses to clinical disease in a minority of cases.

Patients who are infected with HIV are predisposed to reactivation of latent TB infection, and also to a rapid progression of recently acquired infection.

Other predisposing factors may be vitamin D deficiency



Laboratory Testing: Sputum



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Sputum specimens should be relatively **FRESH** (less than 1 day old) to minimise contamination. **PURULENT** specimens are best. Two to three samples of ≥5mL should be collected approximately 8-24 hours apart with at least one from early morning.

Samples taken **EARLY MORNING** (that is, shortly after patient waking) have the greatest yield. When the cough is dry, physiotherapy, postural drainage or inhalation of nebulised saline ('sputum induction') before expectoration may be helpful.

Laboratory Testing: Direct Smear Microscopy





ZN Stain

PA Stain





Auramine-phenol staining is more sensitive than that by the Ziehl-Neelsen method, and is therefore more suitable for assessment of smears from clinical specimens: variable sensitivity & specificity reported across journals – UK recommended method

UK SMI: B 40 Investigation of specimens for Mycobacterium species (publishing.service.gov.uk)

Microscopy result should be reported within one working day of receipt of the specimen

Culture Detection





Molecular Detection





High clinical suspicion of MTb OR PA Smear Positive

Direct Cepheid

Result: 2hrs



Direct inoculation of the cartridge, ultrasonic lysis of the organisms to release DNA

DNA mixed with dry PCR reagents

Realtime Semi-Nested PCR

Rifampicin Resistance gene RpoB detected

The Xpert MTB/RIF assay (Xpert), the first point-of-care assay for tuberculosis (TB), was endorsed by the World Health Organization in December 2010

Local Molecular Testing – Early classification of MTb Vs MDR MTb

Detect presence Detect resistance Determine the species Tests to detect presence of Allows identification to species Purpose of test Tests to detect resistance to M. tuberculosis complex and anti tuberculous drugs e.g. level (includes tuberculous some non-tuberculous (atypical rifampicin & isoniazid mycobacteria). mycobacteria). e.g. in rifampicin resistance, Amplification of target Target is a sequence i.e. How it works specific to the M. tuberculosis mutation of rpoB gene occurs mycobacterial sequences complex (MTBC). in a small area in the gene. This based on species specific can be amplified. This complex includes targets M. tuberculosis, M. bovis, M. africanum and BCG Positive test indicates infection Known resistance mutations Determines the exact species with any of the above present can be detected within the mycobacterial complex and may determine What it means the species if a non-tuberculous strain If the target is a sequence specific to MTBC then other non-tuberculous species will not be detected. Specimen Type Can be used directly on Can be used directly on Can be used directly on specimen or on culture specimen or culture specimen or culture

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PRESENCE RESISTANCE MDR TB



Laboratory Mixed Methods Approach

Figure 2. Microbiology tests available for the diagnosis of active TB





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Microscopy: Fast, Cheap & indicates bacterial load



Molecular Tests: expensive, enable MTB detection before culture



Culture is needed to be able to perform sensitivity testing & whole genome sequencing



Remarkable people. Extraordinary place. Susceptibility Testing





M. tuberculosis is usually detected and identified within 7 to 21 days depending upon biomass, which is also reflected in the smear

result.



Drug susceptibility testing has 3 main goals:

•to facilitate the management of individual patients, particularly if drug resistance is likely

•to provide data on which to plan drug combinations for treatment

•to provide a surrogate measure of the relative effectiveness of tuberculosis control programmes

Results of tests for primary therapeutic agents should be completed within 14 days of receipt



AST mostly performed by WGS Standard AST can take up to 40 days



WGS



WGS based identification service from liquid or solid culture media provided free to the NHS approx. 7d

Drug susceptibility testing and genotypic resistance prediction for M. tuberculosis complex:

Genotypic drug susceptibility predictions are made for all isolates. Routine phenotypic susceptibility testing for first line agents (isoniazid / rifampicin / pyrazinamide / ethambutol) is **no longer performed**.

If WGS predicts resistance or if WGS data is not sufficiently clear to accurately make a prediction, phenotypic testing of first line agents will be performed. Testing for second and third line agents will be performed for multi-drug resistant isolates when clinically indicated (Early information linking XDR cases is vital)

England world leaders in the use of whole genome sequencing to diagnose TB - GOV.UK (www.gov.uk)







Determination of M. tuberculosis isolate relatedness – based on **SNP differences** determined by WGS, provided free to the NHS and for the support of outbreak investigations, detection of laboratory cross-

contamination events

Drug	Mutation	Nucleotides	Support (A/C/G/T)	Source	Prediction
AK	rss_*1484*	G->N	(0/0/4/0)	Line-probe	F
AK	rrs_*1401*	A->N	(2/0/0/0)	Line-probe	F
CAP	rrs_*1402*	C->N	(0/3/0/0)	Line-probe	F
			-		
Drug	Mutation	Nucleotides	Support (A/C/G/T)	Source	Prediction
EMB	embB_D354A	GAC->GCC	(0/0/26/0)	derived-(3/5)	R
			(1/31/0/1) (0/27/0/0)		
INH	katG_S315T	AGC->ACC	(41/0/0/0)	Line-probe/derived-(471/480)	R
			(0/44/0/0) (0/43/0/0)		
KAN	rrs_C517T	C->T	(0/0/0/49)	derived-(1/6)	R
RIF	rpoB_*449*	CTG->CNG	(0/39/0/0)	Line-probe	F
			(3/0/0/32) (0/0/36/0)		

SNPs result from mutations at a single position in the DNA sequence. Because SNPs gradually accumulate over time, the number of SNPs that differ between isolates (SNP distance) can provide information about whether the TB cases could be the result of recent transmission

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NTM



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Non-tuberculous mycobacteria (NTM) are also increasingly encountered as a cause of disease in humans.

Unlike *M. tuberculosis,* isolation of an NTM species from specimens such as sputum does not equate to disease – the microbiology results need to be interpreted in conjunction with clinical and radiological findings.

Rapid Growers



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- Mycobacterium abscessus, Mycobacterium chelonae, Mycobacterium fortuitum
- These grow in routine culture SA bottles
- D3 onwards
- *M. abscessus* more so than the other non-tuberculous mycobacteria are an increasing problem for the cystic fibrosis patient group. Testing should be considered in cystic fibrosis patients who show deteriorating lung function but where no clear pathogen has been identified. The Cystic Fibrosis Trust microbiology standards recommend routine screening of NTM at least once a year for all patients able to produce sputum and all *M. abscessus* positive isolates should be referred to the appropriate reference laboratory for strain typing (VNTR).

Slow Growers



Mycobacterium avium – intracellulare group (MAI)

There are currently three species within the MAC and they are *M. avium*, *M. intracellulare and M. chimaera*. Additionally, there are now three valid named subspecies of *M. avium*: *M. avium* subsp. *avium*; *M. avium* subsp. *avium*; *M. avium* subsp. *paratuberculosis*; and *M. avium* subsp. *Silvaticum*.

M. chimaera, a slow growing NTM found in the environment has been implicated recently in several cases of endocarditis or deep infection following cardiac surgery involving the use of cardiac bypass equipment.

Mycobacterium gordonae Mycobacterium kansasii Mycobacterium malmoense Mycobacterium marinum Mycobacterium ulcerans Mycobacterium xenopi





Once treatment is initiated we repeat cultures until the patient becomes culture & smear negative

Persistent Smear positive occurs when patients have extensive cavities – this can influence length of treatment or instigate a change in treatment.

We will re-refer for WGS derived AST

Culture Detection MALDI-TOF (Near Future)





Resources



https://assets.publishing.service.gov.uk/government/uploads/system/uploads/ /attachment_data/file/489198/Molecular_diagnosis_of_tuberculosis_for_heal thcare_professionals.pdf

https://assets.publishing.service.gov.uk/government/uploads/system/uploads/ /attachment_data/file/684543/B_40i7.2.pdf